

SOP: LFA 00071 SOP For Antimicrobial Effectiveness Testing

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1 Objective

To provide the proper procedure to follow for effective antimicrobial testing using standard culture

2 Scope

Applicable to Microbiology Lab

3 Responsibility

By Microbiologist

4 Accountability

Head of the Department

5 Procedure

- 5.1. Precautions to be observed during testing
 - 5.1.1. Sterilized glasses should be used during testing.
 - 5.1.2. It is important to pre-incubate any media that will be used.
 - 5.1.3. Standard culture to be used should not be more than four passages.
- 5.2. Type of Organism to use
 - 5.2.1. Choose the culture from the following microorganisms:
 - Candida albicans ATCC No. 10231
 - Aspergillus Niger ATCC No. 16404
 - Escherichia coli ATCC No. 6538

The microorganism to be used should not be more than five passages from the original ATCC culture of any of the equivalent cultures.

- 5.3. Preparing the Inoculums
 - 5.3.1. Follow the SOP of Preparation for Culture Suspension to prepare the inoculums.
 - 5.3.2. In order to harvest the bacteria and Candida culture, use a sterile peptone saline. Wash the surface growth and collect in an appropriate glass container. Add enough sterile peptone saline in order to gather a microbial count of approximately 1×10^8 CFU/ml.
 - To harvest Aspergilus niger cells, use a sterile peptone saline with 0.05% polysorbate 80, adding enough sterile peptone saline to achieve a count of 1×10^8 CFU/ml.
 - 5.3.3. Obtain the total number of CFU/ml for each suspension using media conditions and microbial recovery times indicated in the table below in order to confirm the initial CFU/ml. The value would help in calibrating the inoculum size used in the test. Bacterial and yeast suspensions to be used for testing should be used within 24 hours of the harvest; however fungal preparations can be stored for up to 7 days in the fridge.

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5.4. Test Procedures

niger

Table 1: Table for Culture Conditions for Inoculum Preparation:								
				Microbial				
Organism	Sutable	Incubation	Incubation	Recovery				
	Medium	Temperature	Time	Incubation				
				Time				
Escherichia	SCM / SCA	$32.5 \pm 2.5C$	10 to 24 hours	2 to 5 days				
coli	SCM / SCA	02.0 ± 2.00	10 to 24 hours	3 to 5 days				
Pseudomonas	SCM / SCA	$32.5 \pm 2.5C$	10 to 24 hours	3 to 5 days				
aeruginosa	SOM / SOA	32.3 ± 2.30	10 to 24 hours	o to o days				
Staphylococcus	SCM / SCA	$32.5 \pm 2.5C$	10 to 24 hours	3 to 5 days				
aureus	SOM / SOA	32.3 ± 2.30	10 to 24 hours					
Candida	SCM / SCA	$22.5 \pm 2.5C$	44 to 52 hours	3 to 5 days				
albicans	DOM / BOA	22.0 ± 2.0C	44 10 02 110418	o to o days				
Aspergillus	SCM / SCA	$22.5 \pm 2.5C$	6 to 10 days	3 to 7 days				
:		22.0 ± 2.00	l o to to days	j o to i days				

Table 1: Table for Culture Conditions for Inoculum Preparation:

- 5.4.1. The testing can be performed in five containers if there is sufficient product volume available; however, if there is insufficient product, one can also use five sterile capped bacteriological containers where suitable volume can be transferred.
- 5.4.2. Place the prepared and standardized inoculum to each container and mix. The suspension inoculum volume used should be between 0.5% and 1% of the product?s volume.
- 5.4.3. The microorganism concentration volume is such that the final concentration of the test preparation after the inoculation phase should be between 1×10^5 and 1×10^6 CFU/ml of the product.
- 5.4.4. The initial concentration of viable microorganism is estimated based on the microorganism concentration volume added to each of the standardized inoculum as determined by the plate-count method.
- 5.4.5. Incubate the inoculated containers as $22.5\pm2.5^{\circ}$ C. Check the samples at certain intervals as specified in 5.5 and document any changes observed during the intervals.
- 5.4.6. Determine by using the plate-count method the total number of the CFU present in each of the intervals.
- 5.4.7. Add an in-activator or neutralizer of the specified antimicrobial to the

plate count or if required, to the appropriate dilution that is prepared for plating.

5.4.8. To calculate the changes, use the calculated concentration of the CFU/ml at present of the test, and calculate the change in log 10 value of the CFU/ml concentration for each of the microorganism at the specified test interval. Document the changes in the log reductions.

Table 2: Criteria for Frequency and Acceptance

Category of products	Case	Bacterial Log Reduction					Fungal Log Reduction			
products		6	24	2	$7 \mathrm{th}$	14th	28th	$7 \mathrm{th}$	14th	28th
		hrs	hrs	days	day	day	day	day	day	day
Parenteral and	A	2	3	-	-	-	NR	2	-	NI
opthalmic	В	-	1	-	3	-	NI	-	-	NI
preperations										
Topical	A	-	-	2	3	-	NI	-	-	NI
preperations	В	-	-	-	-	3	NI	-	-	NI
Oral preparations	A	-	-	-	-	3	NI	-	-	NI

As Per USP:

CATEGORY	BACTERIA	YEAST/MOLDS			
Injections					
 parenteral injections emulsions otic sterile nasal products ophthalmic products made with aqueous bases/vehicles 	 Not less than 1.0g reduction from the first calculated count at seven days Not less than 3.0 log reduction from the first count at 14 days No increase from the 14 days count at 28 days 	• No increase from the first calculated count at 7, 14, and 28 days			
 Topical products with aqueous base Non-sterile nasal products Emulsions including those applied to the mucous membrane 	 Not less than 2.0 log reduction from the first count at 14 days No increase from the 14 days count at 28 days 	• No increase from the first calculated count at 14 and 28 days			
• Oral products other than antacids (made with aqueous bases/vehicles)	 Not less than 1.0 log reduction from the first count at 14 days No increase from the 14 days count at 28 days 	• No increase from the first calculated count at 14 and 28 days			
• Antacids (made with aqueous base)	• No increase from the first calculated count at 14 and 28 days	• No increase from the first calculated count at 14 and 28 days			

As Per Harmonized Criteria

Category of products	Case	Bacterial Log Reduction						Fungal Log Reduction		
		6	24	2	7th	14th	28th	7th	14th	28th
		hrs	hrs	days	day	day	day	day	day	day
Parenteral and	A	2	3	-	-	-	NR	2	-	NI
opthalmic	В	-	1	-	3	-	NI	-	-	NI
preperations										
Topical	A	-	-	2	3	-	NI	-	-	NI
preperations	В	-	-	-	-	3	NI	-	-	NI
Oral	A	Λ				3	NI			NI
preperations		_	_	_	_	3	111	_	_	111
Antacids	-	-	-	-	-	NI	NI	-	NI	NI

6 Abbreviations

- **SOP** Standard Operating Procedure
- ATCC American Type of Culture Collection
 - CFU Colony Forming Unit
 - **SCM** Soybean Casein Digest Medium
 - $\mathbf{SCA}\,$ Soybean Casein Digest Agar
 - ${\bf SDA}\,$ Sabouraud Dextrose Agar
 - $^{\circ}\,$ Degree Centrigrade
 - $\% \ {\rm Percentage}$
 - ${\bf EP}\,$ European Pharmacopoeia
 - USP United State Pharmacopoeia